

## Enfumafungin Derivative MK-3118 Shows Increased *In Vitro* Potency against Clinical Echinocandin-Resistant *Candida* Species and *Aspergillus* Species Isolates

Cristina Jiménez-Ortigosa, a Padmaja Paderu, a Mary R. Motyl, b David S. Perlina

Public Health Research Institute, New Jersey Medical School, Rutgers, The State University of New Jersey, Newark, New Jersey, USA<sup>a</sup>; Merck, Sharp & Dohme Corp., Kenilworth, New Jersey, USA<sup>b</sup>

MK-3118 is as an orally active new antifungal in the early stage of clinical development that inhibits the biosynthesis of  $\beta$ -(1,3)-glucan. We evaluated the *in vitro* activity of this compound against wild-type and echinocandin-resistant (ER) isolates containing mutations in the *FKS* gene(s) of *Candida* spp. and *Aspergillus* spp. MK-3118 demonstrated enhanced efficacy for most *C. albicans* and *C. glabrata* ER isolates relative to caspofungin, with decreased MICs and half-maximal inhibitory concentrations (IC<sub>50</sub>s).

"he echinocandins are first-line agents for treating severe invasive fungal infections (IFIs) (1), being fungicidal against yeast and fungistatic against molds. They alter the integrity of the fungal cell wall via the inhibition of the synthesis of the  $\beta$ -(1,3)-glucan, its major component (2). Specifically, echinocandins target the catalytic subunit of the enzymatic complex β-(1,3)-glucan synthase, encoded by the FKS genes. Reduced susceptibility to echinocandins is associated with mutations in two specific regions in the FKS genes known as hot spots (HS) 1 and 2 that lead to clinical failure or poor response to the therapy (3). The three echinocandins approved by the Food and Drug Administration (FDA) for the treatment of IFIs (caspofungin, anidulafungin, and micafungin) are available only in intravenous formulation, which limits their use in the treatment of less-severe infections or as oral stepdown agents. Enfumafungin is one among several new fungal triterpenoid glycosides isolated from the fermentation of Hormonema sp. (4) that present potent in vitro antifungal activity by inhibiting the  $\beta$ -(1,3)-glucan synthase (5). Recently, a semisynthetic derivative of enfumafungin, MK-3118 (Fig. 1), which is being evaluated as an oral therapy for fungal infections, was described (6). This new compound showed MIC values of  $\leq 1 \mu g/ml$ and ≤0.015 µg/ml against 160 strains of 7 Candida spp. and 40 Aspergillus spp., respectively (7). Moreover, MK-3118 showed promising in vivo efficacy in murine models of candidiasis and aspergillosis (7, 8). To better understand the antifungal efficacy of MK-3118, we evaluated this new compound against a well-characterized panel of echinocandin-resistant (ER) fks mutants derived from patients who failed echinocandin therapy.

Antifungal susceptibility testing was performed in triplicate for

a collection of 95 Candida strains (20 C. albicans, 20 C. glabrata, 2 C. dubliniensis, 15 C. krusei, 19 C. parapsilosis, and 19 C. tropicalis) that included 30 isolates showing an echinocandin resistance (ER) phenotype (caspofungin [CAS] MIC  $\geq$  0.5 µg/ml) and for a panel of 40 Aspergillus strains (14 A. fumigatus, 10 A. flavus, 10 A. niger, and 6 A. terreus) that included 1 isolate showing an ER phenotype (9) in accordance with the guidelines described in CLSI documents M27-A3 and M38-A2 (10, 11). In the case of the Candida isolates, MICs were also determined in the presence of 50% human serum (Sigma-Aldrich) (from human male blood, type AB) or mouse serum (Millipore) for C. glabrata isolates. C. parapsilosis ATCC 22019 and C. krusei ATCC 6258 were used as quality control strains. Caspofungin and MK-3118 were obtained as standard powders from their manufacturer (Merck & Co. Inc., Rahway, NJ), and stock solutions were prepared by dissolving the compounds in water or 100% dimethyl sulfoxide (MK-3118).

The MIC distributions of the *Candida* isolates after 48 h of growth at 35°C for CAS and MK-3118 are shown in Table 1. MK-3118 did not show significant differences in MIC values for the wild-type (WT) isolate population, although overall it presented

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Address correspondence to David S. Perlin, perlinds@njms.rutgers.edu. Copyright © 2014, American Society for Microbiology. All Rights Reserved. doi:10.1128/AAC.02145-13

FIG 1 Structures of enfumafungin and MK-3118, a semisynthetic enfumafungin derivative.

TABLE 1 MIC distributions of CAS and MK-3118 in the presence or absence of serum for the Candida isolates included in this study

		Caspofungin			MK-3118			
Species	Phenotype (no. of isolates)	Mode value [MIC <sub>50</sub> range (mg/liter)] <sup>a</sup>			Mode value [MIC <sub>50</sub> range (mg/liter)] <sup>a</sup>			
		No serum	50% serum	$Ratio^b$	No serum	50% serum	$Ratio^b$	
C. albicans	WT (10)	≤0.03 (≤0.03–0.06)	0.12 (0.12–1)	4	≤0.03 (≤0.03-0.06)	0.5 (0.5–1)	16	
	ER (10)	2 (≤0.03–4)	≥16 (1-≥16)	8	1 (≤0.03-1)	$0.5 - \ge 16 \ (0.5 - \ge 16)$	2–16	
C. glabrata	WT (9)	$0.06 (\leq 0.03-1)$	0.5 (0.5–2)	8	0.12 (0.12–0.5)	2-4 (2-8)	16–32	
	ER (11)	16 (1–16)	≥16	1	0.5 (0.25–8)	≥16 (4-≥16)	32	
C. dubliniensis	WT (1)	0.06	0.25	4	0.12	1	8	
	ER (1)	0.06	0.5	8	0.12	1	8	
C. krusei	WT (11)	0.12 (0.12–0.5)	2 (2-4)	16	0.5 (0.25–0.5)	8 (8-≥16)	16	
	ER (4)	0.12-8 (0.12-8)	≥16 (2-≥16)	2-133	0.25-2 (0.25-2)	≥16 (1-≥16)	8-64	
C. parapsilopsis	WT (19)	1 (0.12–16)	≥16 (4-≥16)	16	0.5 (0.25–8)	≥16 (2-≥16)	32	
C. tropicalis	WT (15) ER (4)	$\leq 0.03 \ (\leq 0.03 - 0.06)$ 2 ( $\leq 0.03 - 2$ )	0.5 (0.25–0.5) ≥16	16 8	0.12 (0.06–0.25) 0.5 (0.25–4)	2 (1–8) 8-≥16 (4-≥16)	16 16–32	

<sup>&</sup>lt;sup>47</sup> Data represent mode values and MIC ranges after 48 h of growth at 35°C. All values represent averages of the results of triplicate experiments with less than 15% variance.

enhanced in vitro efficacy compared to that of CAS for nearly all echinocandin-resistant isolates, especially among the C. albicans and C. glabrata isolates, where the MIC values decreased by 1- to 8-fold and 4- to 32-fold, respectively. C. tropicalis isolates showed a 4-fold decrease in MIC, while the fold change for C. krusei ER isolates was 2 to 4 times lower. Specifically, 50% of the ER isolates of *C. albicans* showed MIC values for CAS of  $\geq 2$  mg/liter whereas 70% of the ER isolates showed an MIC value of  $\leq$  0.5 mg/liter for MK-3118 after 48 h of growth (4- to 8-fold change). Moreover, only 30% of the ER strains showed MIC values of ≤0.25 mg/liter for CAS, while 60% were below this level for MK-3118. The decrease in the MIC values was genotype dependent. Thus, prominent mutations conferring modification of Ser 645 within hot spot (HS) 1 of Fks1p for *C. albicans* showed a 4- to 16-fold reduction in MIC values whereas strains containing modifications at Phe 641 showed results for CAS and MK-3118 that were similar. In addition, 64% of C. glabrata ER strains showed MIC values of ≤0.5 mg/liter for MK-3118 after 48 h of growth whereas all ER isolates showed MIC values of  $\geq 1$  mg/liter for CAS. The decrease in the MIC values was not genotype dependent in C. glabrata, as mutations in either the FKS1 gene or the FKS2 gene showed comparable results (8- to 32-fold reduction in both cases) (Table 2). Similar results were described by Pfaller and collaborators (12), who found that, in a cohort of wild-type clinical isolates (without FKS mutations), there was little or no difference in MIC values between MK-3118 and CAS by broth microdilution for C. albicans, C. krusei, C. parapsilosis, and C. tropicalis. The only exception was C. glabrata, where MK-3118 was 8-fold more potent than CAS. Moreover, 71% of clinical isolates harboring mutations in the FKS gene(s) were inhibited by MK-3118 at  $\leq 1$  mg/liter (12), which correlates well with our data.

The echinocandins are highly bound to serum proteins, with a rate of 98% reported for caspofungin (13), which alters its antifungal properties. In fact, it has been reported that the addition of 50% of human serum increased caspofungin MICs an average of 2-fold with a range of 1- to 16-fold (14). In order to ascertain if the relative in vitro potency of MK-3118 was affected by serum, 50% (wt/vol) human serum was added to the MIC plates, as previously described (13). In the case of C. glabrata isolates, 50% mouse serum was used because human serum can inhibit the growth of this organism (15). The addition of serum to the plates increased the MIC of MK-3118 an average of 16-fold with a range of 8- to 64-fold, four times higher than the values obtained for CAS (Table 1). The reduced antifungal properties of this compound in the presence of serum suggested that protein binding was having a direct effect on the drug, perhaps by altering its ability to inhibit glucan synthase, as was observed previously for the echinocandins (14, 16).

Abnormal growth morphology was used to establish a minimum effective concentration (MEC) for Aspergillus spp. after 24 h

TABLE 2 MIC distributions of CAS and MK-3118 for the C. albicans and C. glabrata isolates harboring mutations in the FKS genes included in this study

		Mutation(s) found	MIC (mg/liter)		
Strain	Species	Fks1p	Fks2p	CAS	MK-3118
DPL18	C. albicans	F641S		0.5	1
DPL20	C. albicans	S645P		4	0.5
DPL22	C. albicans	S645P/S		0.03	0.06
DPL1007	C. albicans	F641S		1	1
DPL1008	C. albicans	S645P		4	1
DPL1009	C. albicans	S645Y		2	0.12
DPL1010	C. albicans	S645F		2	0.12
DPL1011	C. albicans	S645F + R1361R/H		2	< 0.03
DPL1012	C. albicans	D648Y		0.25	< 0.03
DPL1013	C. albicans	P649H		0.25	0.25
DPL23	C. glabrata		F659del	>16	4.00
DPL26	C. glabrata		F659S	>16	0.50
DPL30	C. glabrata		S663P	>16	0.50
DPL33	C. glabrata		D666E	2.00	0.25
DPL34	C. glabrata		P667T	2.00	0.50
DPL38	C. glabrata	F625S		2.00	4.00
DPL39	C. glabrata	S629P		16.00	0.50
DPL41	C. glabrata	D632G		2.00	0.50
DPL42	C. glabrata	D632G		16.00	8.00
DPL155	C. glabrata		F659V	4.00	2.00
DPL236	C. glabrata		L664R	1.00	0.50

<sup>&</sup>lt;sup>b</sup> Data represent fold change after adding 50% of serum to the MIC plates.

TABLE 3 MEC distributions of CAS and MK-3118 for the *Aspergillus* isolates included in this study

	Phenotype	Mode value and $MEC_{50}$ range $(mg/liter)^a$					
Species	(no. of isolates)	Caspofungin	MK-3118				
A. flavus	WT (10)	0.12 (0.06–2)	8 (2.0–16)				
A. fumigatus	WT (1) ER (1) WT (6 [ITR <sup>s</sup> ]) <sup>c</sup> WT (8 [ITR <sup>r</sup> ]) <sup>b</sup>	0.12 >16 0.12 (0.12–0.25) 0.12 (0.06–0.25)	0.12 0.12 8 (0.12–8) 0.25 (≤0.03–8)				
A. niger A. terreus	WT (10) WT (6)	0.12 (0.06–0.12) 0.06 (0.06–0.25)	0.12 (≤0.03–0.25) 0.12 (0.06–0.12)				

 $<sup>^{\</sup>overline{a}}$  Data represent mode values and MEC ranges after 48h of growth at 35°C. All values represent averages of the results of triplicate experiments with less than 15% variance.

and 48 h of growth at 35°C. MK-3118 and caspofungin were quite active against the four species of filamentous fungi analyzed, including eight *A. fumigatus* strains with an azole-resistant phenotype. Interestingly, the growth of all *Aspergillus* isolates was completely inhibited by treatment with high concentrations (8 to 16  $\mu$ g/ml) of MK-3118. These data are in accord with those of Pfaller et al. (17), who showed that MK-3118 was active against 71 *Aspergillus* isolates, including 8 itraconazole-resistant isolates (MIC  $\geq$  4  $\mu$ g/ml). Since echinocandin-resistant isolates from *Aspergillus* spp. have rarely been observed, MK-3118 was tested against the only ER strain of *A. fumigatus* available to date, which presents the

amino acid substitution S678P, equivalent to that of the S654P of *C. albicans* (9). In this isolate, MK-3118 showed prominent increased potency with an MIC that was 133 times less than that of CAS after 24 h of growth (Table 3).

To better assess direct inhibition of MK-3118 on glucan synthase, the kinetic inhibition parameter IC<sub>50</sub> (half-maximal inhibitory concentration) was determined for glucan synthases from wild-type and fks-containing strains. Product-entrapped 1,3-β-Dglucan synthase complexes (GS) were extracted from wild-type and ER strains containing fks mutations from C. albicans (1 WT and 3 fks mutant strains), C. glabrata (1 WT and 2 fks mutant strains), and A. fumigatus (1 WT strain and 1 fks mutant strain), as described previously (18, 19). As expected, evaluation of kinetic inhibition of product-entrapped enzymes isolated from ER strains yielded lower IC<sub>50</sub>s for MK-3118 than for CAS in Candida albicans (3- to 5.5-fold) and Candida glabrata (3.5- to 62-fold) (Fig. 2A and Table 4). An exception was observed for glucan synthase harboring the mutation F641S, as inhibition of GS activity did not exhibit any variation in the percentage of incorporation even after exposure to high doses of the drugs (10 μg/ml); it was not possible to obtain an in-range  $IC_{50}$  for MK-3118 (Fig. 2A). In the case of A. fumigatus, a decrease of 28-fold was detected in the IC<sub>50</sub> of a prominent ER strain (Fks1p-S678P) compared to that of the WT for MK-3118 (Fig. 2B and Table 4), indicating a potential advantage of MK-3118 over echinocandin drugs for certain echinocandin-resistant strains.

In summary, MK-3118 was highly active on most fks-mediated echinocandin-resistant strains, especially those from *C. albicans* and *C. glabrata*. It was also active on *Aspergillus* spp. at high con-

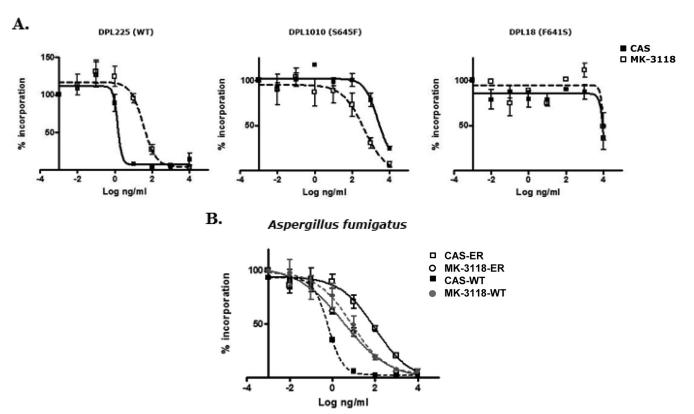


FIG 2 Antifungal inhibition profiles for product-entrapped 1,3- $\beta$ -glucan synthase enzyme complexes (GS) for caspofungin (CAS) and MK-3118 for wild-type and ER clinical isolates. GS inhibition was assessed by the incorporation of [ $^3$ H]glucose into radiolabeled product. (A) *Candida albicans*. (B) *Aspergillus fumigatus*.

<sup>&</sup>lt;sup>b</sup> Isolates with a WT FKS1 gene and sensitive to azoles. ITR, itraconazole.

<sup>&</sup>lt;sup>c</sup> Isolates with a WT FKS1 gene but resistant to azoles.

TABLE 4 *In vitro* whole-cell susceptibility and 1,3- $\beta$ -glucan synthase inhibition profiles of caspofungin and MK-3118 for representative strains included in the study<sup>a</sup>

				$MIC (mg/liter)^b$					
		Fksp phenotype		CAS		MK-3118		IC <sub>50</sub> (ng/ml)	
Strain	Organism	Fks1p	Fks2p	No serum	50% serum	No serum	50% serum	CSF	MK-3118
DPL225	C. albicans	WT		< 0.03	0.12	0.03	0.5	1.421	32.95
DPL1010	C. albicans	S645F		2	>16	0.12	1	2,321	423.1
DPL1012	C. albicans	D648Y		0.25	16	< 0.03	2	149.8	51.58
DPL18	C. albicans	F641S		0.5	>16	1	16	$>2,500^{c}$	$>2,500^{c}$
DPL1021	C. glabrata	WT	WT	0.06	1	0.25	4	125.9	108.1
DPL235	C. glabrata	WT	F659V	>16	>16	0.5	8	1,945	542.1
DPL32	C. glabrata	WT	D666G	2	>16	0.5	4	2,273	36.25
DPL1034	A. fumigatus	WT		0.12	ND	0.12	ND	0.65	7.81
DPL1035	A. fumigatus	$S678P^d$		>16	ND	0.12	ND	100.4	3.58

<sup>&</sup>quot; MIC values represent whole-cell susceptibility; IC<sub>50</sub> values represent 1,3-β-glucan synthase inhibition. CSF, cerebrospinal fluid; ND, not determined.

centrations of the drug and was active against a highly ER strain. As observed previously with echinocandin drugs, serum shifted the relative efficacy of the new compound MK-3118, which was most effective against echinocandin-resistant strains.

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## **REFERENCES**

- Pappas PG, Kauffman CA, Andes D, Benjamin DK, Jr, Calandra TF, Edwards JE, Jr, Filler SG, Fisher JF, Kullberg BJ, Ostrosky-Zeichner L, Reboli AC, Rex JH, Walsh TJ, Sobel JD; Infectious Diseases Society of America. 2009. Clinical practice guidelines for the management of candidiasis: 2009 update by the Infectious Diseases Society of America. Clin. Infect. Dis. 48:503–535. http://dx.doi.org/10.1086/596757.
- Denning DW. 2003. Echinocandin antifungal drugs. Lancet 362:1142– 1151. http://dx.doi.org/10.1016/S0140-6736(03)14472-8.
- Perlin DS. 2007. Resistance to echinocandin-class antifungal drugs. Drug Resist. Updat. 10:121–130. http://dx.doi.org/10.1016/j.drup.2007.04.002.
- Onishi J, Meinz M, Thompson J, Curotto J, Dreikorn S, Rosenbach M, Douglas C, Abruzzo G, Flattery A, Kong L, Cabello A, Vicente F, Pelaez F, Diez MT, Martin I, Bills G, Giacobbe R, Dombrowski A, Schwartz R, Morris S, Harris G, Tsipouras A, Wilson K, Kurtz MB. 2000. Discovery of novel antifungal (1,3)-beta-D-glucan synthase inhibitors. Antimicrob. Agents Chemother. 44:368–377. http://dx.doi.org/10.1128/AAC.44.2.368 -377.2000.
- 5. Peel M, Fan W, Mamai A, Hong J, Orr M, Ouvry G, Perrey D, Liu H, Jones M, Nelson K, Ogbu C, Lee S, Li K, Kirwan R, Noe A, Sligar J, Martensen P, Balkove J, Greenlee CM, Meng D, Parker D, Wildonger K, Liberator P, Abruzzo G, Flattery A, Galgoci A, Giacobbe R, Gill C, Hsu MJ, Misura A, Nielsen J, Powles M, Racine F, Dragovic J, Habulihaz B, Balkovec J. 2010. Enfumafungin derivatives: orally active glucan synthase inhibitors, abstr F1-845. Abstr. Intersci. Conf. Antimicrob. Agents Chemother., Boston, MA.
- 6. Motyl MR, Tan C, Liberator P, Giacobbe R, Racine F, Hsu MJ, Nielsen-Kahn, Bowman JJ, Douglas C, Hammond M, Balkovec JM, Greenlee ML, Meng D, Parker D, Peel M, Fan W, Mamai A, Hong J, Orr M, Ouvry G, Perrey D, Liu H, Jones M, Nelson K, Ogbu C, Lee S, Li K, Kirwan R, Noe A, Sligar J, Martensen P. 2010. Abstr. Intersci. Conf. Antimicrob. Agents Chemother., Boston, MA, abstr F1-847.
- Flattery A, Abruzzo G, Gill C, Powles M, Misura A, Galgoci A, Colwell L, Dragovic J, Tong X, Wolff M, Liberator P. 2010. Abstr. Intersci. Conf. Antimicrob. Agents Chemother., Boston, MA, abstr F1-848.
- Flattery A, Abruzzo G, Gill C, Powles M, Misura A, Galgoci A, Douglas C, Hawkins J, Galuska S, Pereira T, Tong S, Wolff M, Song Q, Liberator P. 2010. Abstr. Intersci. Conf. Antimicrob. Agents Chemother., Boston, MA, abstr F1-849

- Rocha EM, Garcia-Effron G, Park S, Perlin DS. 2007. A Ser678Pro substitution in Fks1p confers resistance to echinocandin drugs in Aspergillus fumigatus. Antimicrob. Agents Chemother. 51:4174–4176. http://dx.doi.org/10.1128/AAC.00917-07.
- National Committee for Clinical Laboratory Standards. 2008. Reference method for broth dilution antifungal susceptibility testing of yeasts; approved standard. Clinical and Laboratory Standards Institute document M27-A3, 3rd ed, vol 28. National Committee for Clinical Laboratory Standards, Wayne, PA.
- 11. National Committee for Clinical Laboratory Standards. 2008. Reference method for broth dilution antifungal susceptibility testing of filamentous fungi; approved standard. Clinical and Laboratory Standards Institute document M38-A2, 2nd ed, vol 28. National Committee for Clinical Laboratory Standards, Wayne, PA.
- 12. Pfaller MA, Messer SA, Motyl MR, Jones RN, Castanheira M. 2013. Activity of MK-3118, a new oral glucan synthase inhibitor, tested against Candida spp. by two international methods (CLSI and EUCAST). J. Antimicrob. Chemother. 68:858–863. http://dx.doi.org/10.1093/jac/dks466.
- 13. Hajdu R, Thompson R, Sundelof JG, Pelak BA, Bouffard FA, Dropinski JF, Kropp H. 1997. Preliminary animal pharmacokinetics of the parenteral antifungal agent MK-0991 (L-743,872). Antimicrob. Agents Chemother. 41:2339–2344.
- Paderu P, Garcia-Effron G, Balashov S, Delmas G, Park S, Perlin DS. 2007. Serum differentially alters the antifungal properties of echinocandin drugs. Antimicrob. Agents Chemother. 51:2253–2256. http://dx.doi.org /10.1128/AAC.01536-06.
- Garcia-Effron G, Park S, Perlin DS. 2011. Improved detection of Candida sp. fks hot spot mutants by using the method of the CLSI M27-A3 document with the addition of bovine serum albumin. Antimicrob. Agents Chemother. 55:2245–2255. http://dx.doi.org/10.1128 /AAC.01350-10.
- Odabasi Z, Paetznick V, Rex JH, Ostrosky-Zeichner L. 2007. Effects of serum on in vitro susceptibility testing of echinocandins. Antimicrob. Agents Chemother. 51:4214–4216. http://dx.doi.org/10.1128/AAC.01589-06.
- Pfaller MA, Messer SA, Motyl MR, Jones RN, Castanheira M. 2013. In vitro activitity of a new oral glucan synthase inhibitor (MK-3118) tested against Aspergillus spp. by CLSI and EUCAST broth microdilution methods. Antimicrob. Agents Chemother. 57:1065–1068. http://dx.doi.org/10 .1128/AAC.01588-12.
- Garcia-Effron G, Lee S, Park S, Cleary JD, Perlin DS. 2009. Effect of Candida glabrata FKS1 and FKS2 mutations on echinocandin sensitivity and kinetics of 1,3-beta-D-glucan synthase: implication for the existing susceptibility breakpoint. Antimicrob. Agents Chemother. 53:3690–3699. http://dx.doi.org/10.1128/AAC.00443-09.
- Garcia-Effron G, Park S, Perlin DS. 2009. Correlating echinocandin MIC and kinetic inhibition of fks1 mutant glucan synthases for Candida albicans: implications for interpretive breakpoints. Antimicrob. Agents Chemother. 53:112–122. http://dx.doi.org/10.1128/AAC.01162-08.

<sup>&</sup>lt;sup>b</sup> MEC in the case of A. fumigatus.

<sup>&</sup>lt;sup>c</sup> No significant inhibition at all levels.

<sup>&</sup>lt;sup>d</sup> This mutation is equivalent to S645P in C. albicans (9).